

Stereoselective synthesis of 2,6-dideoxy; 3,6-dideoxy; 2,3,6-trideoxy-inositol 1,4,5-trisphosphate and 6-deoxy-inositol 1,3,4,5-tetrakisphosphate analogues from 6-deoxy-D-inositol precursors

Aloísio Antonio Alves Benício,^a Adilson David Da Silva,^{a,*} Mauro Vieira De Almeida,^a Maria Mirtes Da Silva^b and Stéphane Dov Gero^b

^aDepartamento de Química, Instituto de Ciências Exatas, Universidade Federal de Juiz de Fora, 36036-330 Juiz de Fora MG, Brazil ^bInstitut de Chimie des Substances Naturelles, C.N.R.S., 91198 Gif-sur-Yvette Cedex, France

This article is dedicated to the memory of Stéphane Dov Gero, who passed away on 16 November 1998

Received 28 April 2000; accepted 21 November 2000

Abstract—The novel 2,6-dideoxy-2,2-difluoro; 2,6-dideoxy-2-fluoro; and 2,3,6-trideoxy-2,3-difluoro-Ins(1,4,5)tris(dibutyl)phosphates and 6-deoxy-D-*chiro*-inositol 1,3,4,5-tetrakis(dibutyl)phosphate analogues were synthesized in optically pure form. Modification of Ins(1,4,5)P₃ and Ins(1,3,4,5)P₄ analogues by lipophilic substituents has been investigated in order to produce neutral phosphate derivatives aimed to be incorporated in cell membrane for in vivo evaluation. © 2001 Elsevier Science Ltd. All rights reserved.

It is conceivable that inhibitors of the key enzymes of the phosphoinositide cascade, as phosphatases or kinases, could be of medicinal interest. For example, intracellular signaling pathways mediating the effects of oncogenes on cell growth and transformation offer novel targets for the development of anticancer drugs.¹

As a result of the stimulation of cell surface receptors by a variety of ligands, the membrane-located phosphatidylinositol 4,5-bisphosphate is hydrolyzed by phospholipase C to 1D-*myo*-inositol 1,4,5-trisphosphate [Ins(1,4,5)P₃] and diacylglycerol. The further metabolism of Ins(1,4,5)P₃ is quite complex and involves the action of various kinases and phosphatases. An important initial transformation is the phosphorylation of Ins(1,4,5)P₃ through the action of an ATP-dependent 3-kinase to give 1D-*myo*-inositol 1,3,4,5-tetrakisphosphate [Ins(1,3,4,5)P₄]. While the role of this metabolite is still under study, the suggestion has been made that it may play a role in refilling the intracellular Ca²⁺.²

A major challenge is now the elucidation of the structural basis for interaction of $Ins(1,4,5)P_3$ both with its receptor and with the metabolic enzymes $Ins(1,4,5)P_3$ 3-kinase and

5-phosphatase, and the rational chemical design of agonists, antagonists and enzyme inhibitors.

Several inositol ring-modified and phosphate-modified analogues have been synthesized and progress has been made in understanding the role of phosphate and hydroxyl groups in determining activity of second messengers.³

The D-3-substituted *myo*-inositol analogues are selective inhibitors of the growth of v-*sis*-transformed NIH 3T3 cells.⁴ Furthermore the 6-deoxy $Ins(1,4,5)P_3$ has already been shown to be a full agonist for Ca²⁺ release in permeabilized SH-SY5Y human neuroblastoma cells, a relatively potent $Ins(1,4,5)P_3$ 5-phosphatase inhibitor and a weak substrate for $Ins(1,4,5)P_3$ 3-kinase.⁵

The optically active starting material, 6-deoxy Ins(1,5) diol **1**, was prepared from D-galactose (Scheme 1) according to our earlier developed methodology, using Ferrier rearrangement in the stereoselective sugar to cyclitol transformation step.⁶

1. Results and discussion

In view of the inhibition of the 3-kinase and 5-phosphatase by 2-, 3- or 6-deoxy substituted $Ins(1,4,5)P_3$ derivatives and after the success accounted on the transmembrane incorporation of lipophilic derivatives of $Ins(1,4,5)P_3$ ³ the

Keywords: inositol 1,4,5-trisphosphate; phosphoinositide cascade; lipophilic substituents.

^{*} Corresponding author. Fax: +(32)-3229-3314;

e-mail: david@quimica.ufjf.br



Scheme 1.

preparation of the 2-, 3- and 6-deoxy $Ins(1,4,5)P_3$ derivatives in more lipophilic forms seemed attractive. Additionally, the fluorine atom is a suitable isostere for a hydroxy group. Such an isosteric replacement was viewed as giving rise to a compound which in terms of its physiochemical properties should still be close enough to inositol to enter into the cell via a carrier-mediated process. Furthermore in the last decade a number of deoxy-fluoro-*myo*-Ins(1,4,5)P_3 and deoxy-fluoro-*scyllo*-Ins(1,4,5)P_3 analogues have shown interesting biological results.⁷

Keeping these considerations in mind, we justified the synthesis of 2,6-dideoxy-2,2-difluoro-Ins(1,4,5)tris(dibutyl)-phosphate **9**, 2,6-dideoxy-2-fluoro-Ins(1,4,5)tris(dibutyl)-phosphate **17** and 2,3,6-trideoxy-2,3-difluoro-Ins(1,4,5)-tris(dibutyl)phosphate **14** and 6-deoxy-D-*chiro*-Ins(1,3,4,5)-tetrakis(dibutyl) phosphate **22**.

Compound 1 was treated with benzyl bromide and sodium hydride in DMF to give the tri-O-benzyl derivative 2 in 93% yield (Scheme 2). Hydrolysis of the cyclohexylidene ketal, under mildly acidic conditions, afforded the protected deoxy *myo*-inositol 3, in 95% yield, which presented two free hydroxyls at the 2 and 3 positions. The benzoylation of diol 3 was carried out, in 96% yield, by treatment with benzoyl chloride in pyridine to give the intermediate 4, which was oxidized in the presence of TPAP/NMO to give the 2-inosose 5 in 80% yield. Conversion of this ketone into the 2,2-difluoro derivative 6 was accomplished in 62% yield using (diethylamino)sulfur trifluoride (DAST). The hydrogenolysis of the protected 2,2-difluoro 6, in the

presence of Pd/C 10%, produced the 3-*O*-benzoyl-2,6dideoxy-2,2-difluoro-*myo*-inositol **7**. The latter compound was phosphorylated by reaction with bisbutyloxy(diisopropylamino)phosphine and tetrazole, followed by oxidation with *t*-BuOOH, leading to the protected 2,2difluoro-tris(dibutyl)phosphate **8** in 55% yield. Saponification of the ester intermediate **8**, under basic medium in dichloromethane, afforded the 2,2-difluoro-tris(dibutyl)phosphate **9** in quantitative yield.

In order to produce the 2,3,6-trideoxy-2,3-difluoro-Ins(1,4,5)tris(dibutyl)phosphate **14** the following methods were adopted (Scheme 3). The alcohol **4** was treated with the (diethylamino)sulfurtrifluoride (DAST) with inversion of configuration to yield the 3-*O*-benzoyl-1,4,5-tri-*O*-benzyl-2,6-dideoxy-2-fluoro-*scyllo*-inositol **10**, in 90% yield. Removal of the benzoyl group on **10** under basic conditions gave **11**, in 76% yield, which was fluorinated with DAST, with inversion of configuration to yield the 2,3-difluoro **12** in 75% yield. Hydrogenolysis of the benzyl groups afforded the 2,3-difluoro **13** in 55% yield, which was allowed to be phosphorylated in the presence of bisbutyl-oxy(diisopropylamino)phosphine and tetrazole, followed by oxidation with *t*-BuOOH, leading to **14** in 55% yield.

The deprotection of the intermediate **10** was achieved by hydrogenolysis of benzyl groups to give **15** in quantitative yield (Scheme 4). The 2,6-dideoxy-2-fluoro-Ins(1,4,5)tris-(dibutyl)phosphate **16** was prepared from the triol **15**, in 73% yield, by the same phosphorylation procedure described above. Cleavage of benzoyl group of **16** was





16

Scheme 3.



accomplished in 95% yield by treatment with sodium *n*-butoxide in dichloromethane.

10

15

As described recently,⁸ the protected 6-deoxy Ins(3,4)diol **19** is easily available from cyclohexanone α -hydroxyl (Scheme 1). Saponification of the diester intermediate **19** (Scheme 5), under basic medium, afforded the tetrol **20** in 86% yield, which was allowed to be phosphorylated in the presence of bisbutyloxy(diisopropylamino)phosphine and tetrazole, followed by oxidation with *t*-BuOOH, leading to the tetrakis(dibutyl)phosphate **21** in 55% yield. Hydrogenolysis, in the presence of Pd/C 10% in AcOEt, of intermediate **21** furnished the lipophilic tetrakis(dibutyl)phosphate **22** in 85% yield.

2. Conclusion

17

We have illustrated the potentiality of deoxy cyclitol precursors, stereoselectively produced from D-galactose,^{8,9} to be used for the synthesis of optically pure 2,6-dideoxy-2,2-difluoro; 2,6-dideoxy-2-fluoro and 2,3,6-trideoxy-2,3-difluoro-Ins(1,4,5)tris(dibutyl)phosphates and 6-deoxy-D-*chiro*-inositol 1,3,4,5-tetrakis(dibutyl)phosphate analogues. The 6-deoxy analogues of the well-known second messenger Ins(1,4,5)P₃, possessing a promising biological effect on permeabilized cells,³ have been transformed into lipophilic forms in order to assume their incorporation through the membrane of intact cells.



Preliminary success encountered in vivo in the incorporation of lipophilic analogues of $Ins(1,4,5)P_3$ into cell membrane encouraged the investigation of lipophilic 2,6-dideoxy, 3,6-dideoxy, 2,3,6-trideoxy $Ins(1,4,5)P_3$ and 6-deoxy $Ins(1,3,4,5)P_4$ analogues on Ca^{2+} mobilization.

3. Experimental

3.1. 1,4,5-Tri-*O*-benzyl-2,3-*O*-cyclohexylidene-6-deoxy-D-*myo*-inositol 2

NaH (250 mg, 10.4 mmol) was added, under argon, to diol **1** (1.14 g, 2.22 mmol) dissolved in *N*,*N*-dimethylformamide (15 mL). The mixture was stirred for 10 min before benzylbromide (2.46 mL, 10 mmol) was added. The solution was stirred for 12 h then methanol was added. The reaction mixture was extracted and the organic layer concentrated to dryness. Flash chromatography on silica gel (eluent: AcOEt/heptane) gave the *myo*-inositol **2** (93%). [α]_D= -39° (*c* 0.27; CH₂Cl₂); ¹H NMR (250 MHz; CDCl₃) δ : 4.3 (t, 1H, H-2, $J_{2-1}=J_{2-3}=3$); 4.1 (dd, 1H, H-3, $J_{3-4}=12$); 3.6 (m, 2H, H-4, H-1); 3.3 (m, 1H, H-5); 2.1 (m, 1H, H-6eq); 1.9 (q, 1H, H-6ax, $J_{6ax-1}=J_{6ax-5}=J_{6ax-6eq}=12$); ¹³C NMR (62.5 MHz; CDCl₃) δ : 85 (C-4); 80 (C-3); 77 (C-5); 74 (C-2); 72 (C-1); 30 (C-6); (Found C, 76.86; H, 7.52; C₃₃H₃₈O₅ requires C, 77.01; H, 7.44%).

3.2. 1,4,5-Tri-O-benzyl-6-deoxy-D-myo-inositol 3

Methanol (10 mL, 0.23 mol) and HCl (0.6 mL, 0.02 mmol) were added to the inositol **2** (1.0 g, 1.9 mmol). The solution was stirred at 60°C for 15 h. The reaction mixture was extracted and the organic layer concentrated to dryness. The residue was purified by flash chromatography on silica gel (eluent: AcOEt/heptane) to give the inositol **3** (98%). $[\alpha]_D=4.9^\circ$ (*c* 1.0; CHCl₃); ¹H NMR (300 MHz; CDCl₃) δ : 4.7 (d, 1H, H-3, $J_{2-3}=0$, $J_{3-4}=9$); 4.2 (sl, 1H, H-2); 3.8 (t, 1H, H-4, $J_{4-5}=9$); 3.5 (m, 2H, H-1, H-5); 2.2 (dt, 1H, H-6eq, $J_{6eq-5}=J_{6eq-1}=3$, $J_{6eq-6ax}=12$); 2.0 (q, 1H, H-6ax, $J_{6ax-5}=J_{6ax-1}=J_{6ax-6eq}=12$); ¹³C NMR (75.50 MHz, CDCl₃) δ : 82 (C-4); 78 (C-5); 75 (C-2); 74 (C-3); 70 (C-1); 30 (C-6); (Found C, 74.24; H, 7.01; C₂₇H₃O₅ requires C, 74.63; H, 7.00%)

3.3. 1,4,5-Tri-O-benzyl-3-O-benzoyl-6-deoxy-D-myoinositol 4

To a solution of diol **3** (725 mg, 1.6 mmol) in pyridine (10 mL) was added benzoyl chloride (0.3 mL, 2.10 mmol). The solution was stirred at 10°C for 20 min, extracted with CH₂Cl₂ (2×50 mL) and the organic layer was evaporated under reduced pressure. The residue was purified by flash chromatography on silica gel (eluent: AcOEt/heptane) to afford the benzoylated inositol **4** (80%). $[\alpha]_D=47.0^{\circ}$ (*c* 1.0; CHCl₃); mp 135–136°C; ¹H NMR (300 MHz; CDCl₃) δ : 5.1 (d, 1H, H-3, $J_{3-4}=10$); 4.5 (sl, 1H, H-2); 4.2 (t, 1H, H-4, $J_{4-5}=10$); 3.6 (m, 2H, H-1, H-5); 2.3 (m, 1H, H-6eq); 2.1 (q, 1H, H-6ax, $J_{6ax-5}=J_{6ax-6eq}=J_{6ax-1}=12$); ¹³C NMR (75.50 MHz; CDCl₃) δ : 80 (C-4); 78 (C-5); 75 (C-3); 74 (C-1); 69 (C-2); 30 (C-6); (Found C, 73.56; H, 6.41; C₃₄H₃₄O₆ requires C, 73.36; H, 6.19%).

3.4. 3,4,6-Tri-*O*-benzyl-2-*O*-benzoyl-1-one-5-deoxy-Dmyo-inositol 5

N-oxy-4-methylmorpholine (67.25 mg, 0.57 mmol), tetra-*n*propylammonium perruthenate (13 mg, 0.04 mmol) and molecular sieves (200 mg) were added to a solution of 4 (200 mg, 0.38 mmol) in anhydrous dichloromethane (4 mL). The mixture was stirred for 12 h before addition of isopropanol (1 mL). The stirring was maintained for 30 min before concentration and filtration through silica gel (eluent: AcOEt/heptane). The filtrate was concentrated under reduced pressure and used without further purification (80%); $[\alpha]_{D}=14.1^{\circ}$ (*c* 1.0; CHCl₃); mp 122–125°C; IR 1750 cm⁻¹. ¹H NMR (300 MHz; CDCl₃) δ : 5.5 (d, 1H, H-2, $J_{2-3}=10$); 4.2 (dd, 1H, H-6, $J_{6-5eq}=4$, $J_{6-5ax}=10$); 3.9 (m, 2H, H-4, H-3); 2.6 (m, 1H, H-5eq); 1.8 (q, 1H, H-5ax, $J_{5ax-4} = J_{5ax-5eq} = 10$; ¹³C NMR (75.50 MHz; CDCl₃) δ : 199 (C-1); 82 (C-4); 80 (C-6); 78 (C-2); 75 (C-3); 34 (C-5); (Found C, 76.36; H, 6.17; C₃₄H₃₂O₆ requires C, 76.10; H, 6.01%).

3.5. 1,4,5-Tri-*O*-benzyl-3-*O*-benzoyl-2,2-difluoro-6-deoxy-D-*myo*-inositol 6

Diethylaminosulfur trifluoride (0.3 mL, 2 mmol) was added to a solution of the ketone **5** (500 mg, 0.9 mmol) in anhydrous dichloromethane (5 mL). The solution was stirred for 20 h at room temperature before addition of methanol (5 mL). The reaction mixture was extracted and the organic layer concentrated to dryness. Flash chromatography on silica gel (eluent: AcOEt/heptane) gave the difluoro **6** (62%). $[\alpha]_D=22.3^{\circ}$ (*c* 1.0; CHCl₃); mp 60–62°C; ¹H NMR (200 MHz; CDCl₃) δ : 5.4 (m, 1H, H-3); 3.8 (t, 1H, H-4, $J_{4-5}=J_{3-4}=9$); 3.7–3.5 (m, 2H, H-1, H-5); 2.6 (m, 1H, H-6eq); 1.8 (q, 1H, H-6ax, $J_{6ax-1}=J_{6ax-6eq}=J_{6ax-5}=12$); ¹³C NMR (50 MHz, CDCl₃) δ : 123–120–116 (C-2, $J_{2-F}=252$); 81 (C-4, $J_{4-F}=7$); 76 (C-5); 73 (C-1, $J_{1-F}=19$); 71 (C-3, $J_{3-F}=19$); 32 (C-6, $J_{6-F}=7$); (Found C, 72.17; H, 6.02; C₃₄H₃₂F₂ requires C, 71.94; H, 5.86%).

3.6. 3-O-Benzoyl-2,6-dideoxy-2,2-difluoro-D-myoinositol 7

Difluoro **6** (250 mg, 0.4 mmol) was dissolved in the minimum amount of AcOEt and hydrogenated for 2 h under 3 psi, in the presence of Pd/C 10% (36 mg). The catalyst was removed by filtration on silica gel (eluent: AcOEt) and the solvent evaporated to afford the desired compound **7** in 93% yield. $[\alpha]_{\rm D}=9.2^{\circ}$ (*c* 0.6; CH₃OH); mp 194–196°C; ¹H NMR (300 MHz; CD₃OD) δ : 5.2 (dd, 1H, H-3, $J_{3-4}=10$); 4.5–4.3 (dt, 1H, H-2, $J_{2-F}=52$); 3.9 (m, 1H, H-5); 3.6 (m, 1H, H-1); 3.5 (t, 1H, H-4, $J_{4-5}=10$); 2.2 (m, 1H, H-6eq); 0.8 (q, 1H, H-6ax, $J_{6ax-1}=J_{6ax-6eq}=12$); ¹³C NMR (75.50 MHz; CD₃OD) δ : 169 (C=O); 136–130 (Ph); 122–119–115 (C-2, $J_{2-F}=224$); 77 (C-4); 75 (C-3); 71 (C-5); 69 (C-1); 32 (C-6); (Found C, 54.31; H, 5.05; C₁₃H₁₄O₅F₂ requires C, 54.19; H, 4.90%).

3.7. 3-O-Benzoyl-2,6-dideoxy-2,2-difluoro-D-myoinositol-1,4,5-tris(dibutyl)phosphate 8

To difluoro **7** (86 mg, 0.3 mmol) was added dibutoxy(diisopropylamino)phosphine (480 mg, 1.7 mmol) and the mixture was dried for 0.5 h under vacuum (0.05 mmHg). Sublimated tetrazole (389 mg, 5.55 mmol), dissolved in dry acetonitrile (10 mL) was added under argon to the mixture. The solution was stirred under argon at room temperature for 3 h. The solution was diluted with CH₂Cl₂ before t-BuOOH (1.07 mL, 11.12 mmol) was added. The solution was stirred under argon at room temperature for 12 h. Aqueous sodium thiosulfate was added for neutralization. The product was separated by chromatography on silica gel (eluent: AcOEt/heptane 8:2) (73%). $[\alpha]_{\rm D}=14.6^{\circ} (c$ 1.0; CHCl₃); mp 62–65°C; ¹H NMR (300 MHz; CDCl₃) δ: 5.5 (m, 1H, H-3); 4.8 (m, 2H, H-1, H-2); 4.4 (m, 1H, H-4); 4.1 (m, 12H, OCH2CH2CH2CH3); 2.7 (q, 1H, H-6eq, $J_{6eq-5} = J_{6eq-1} = J_{6ax-6eq} = 8$; 2.1 (m, 1H, H-6ax); 1.7 (m, 12H, OCH₂CH₂CH₂CH₃); 1.4 (m, 12H, OCH₂CH₂CH₂CH₃); 0.9 (m, 18H, OCH₂CH₂CH₂CH₃); ¹³C NMR (75.50 MHz; CDCl₃) δ : 120–117–114 (C-2, $J_{2-F}=201$); 79 (C-5); 75 (C-4); 72 (C-1); 69 (C-3); 68 (OCH₂CH₂CH₂CH₃); 34 (C-6); 32 ($OCH_2CH_2CH_2CH_3$); 19 ($OCH_2CH_2CH_2CH_3$); 13 (OCH₂CH₂CH₂CH₃); (Found C, 50.83; H, 7.41; C₃₇H₆₅O₁₄P₃F₂ requires C, 50.83; H, 7.61%).

3.8. 2,6-Dideoxy-2,2-difluoro-D-*myo*-inositol-1,4,5-tris(dibutyl)-phosphate 9

A solution of sodium butoxide (0.1 mol/L) in dichloromethane was added to compound 8 (86 mg, 0.1 mmol) and the mixture was stirred for 6 h. The reaction mixture was extracted and the organic layer concentrated to dryness. The residue was purified by flash chromatography on silica gel (eluent: AcOEt/heptane 8:2) to furnish the triphosphate 9 (67%). $[\alpha]_{\rm D}=0.44^{\circ}$ (c 2.5; CHCl₃); ¹H NMR (300 MHz; CDCl₃) δ: 4.4 (m, 3H, H-1, H-4, H-5); 4.1 (m 12H, OCH₂CH₂CH₂CH₃); 3.7 (m, 1H, H-3); 2.6 (m, 1H, H-6eq); 2.1 (m, 1H, H-6ax); 1.7 (m, 12H, OCH₂CH₂CH₂CH₃); 1.4 (m, 12H, OCH₂CH₂CH₂CH₃); 0.9 (t, 18H, OCH₂CH₂CH₂CH₃); ¹³C NMR (62.50 MHz, CDCl₃) δ: 80 (C-1); 71-70 (C-3, C-4, C-5); 68 (OCH₂CH₂CH₂CH₂CH₃); 33-32 (OCH₂CH₂CH₂CH₃); 30 (C-6); 19 (OCH₂CH₂CH₂CH₃); 14 (OCH₂CH₂CH₂CH₃); (Found C, 47.38; H, 8.16; C₃₀H₆₁O₁₃P₃F₂ requires C, 47.36; H, 8.08%).

3.9. 1,4,5-Tri-*O*-benzyl-3-*O*-benzoyl-2,6-dideoxy-2-fluoro-*scyllo*-inositol 10

A solution of myo-inositol 4 (1.22 g, 2.26 mmol) in anhydrous dichloromethane (25 mL) was cooled to 0°C before addition of diethylaminosulfur trifluoride (0.6 mL,4.5 mmol) under argon. The solution was stirred for 2 h under argon at room temperature before addition of methanol (10 mL). The mixture was extracted and the organic layer concentrated to dryness. The residue was chromatographed on silica gel (eluent: AcOEt/heptane) to give the monofluoro **10** (90%). $[\alpha]_D = 0.8^\circ$ (*c* 1.0; CHCl₃); mp 94– 95°C; ¹H NMR (300 MHz; C_6D_6) δ : 5.6 (dd, 1H, H-3, $J_{2-3}=9$, $J_{3-4}=12$); 4.5 (m, 1H, H-2, $J_{2-F}=50$); 3.3 (t, 1H, H-4, $J_{4-5}=12$); 3.1 (m, 2H, H-1, H-5); 2.0 (m, 1H, H-6eq); 1.3 (q, 1H, H-6ax, $J_{6ax-6eq}=J_{6ax-1}=J_{6ax-5}=12$); ¹³C NMR (75.50 MHz; C₆D₆) δ : 92 (C-2, $J_{2-F}=187$); 81 (C-4, $J_{4-F}=7$); 73 (C-5); 72 (C-1, $J_{1-F}=19$); 71 (C-3, $J_{3-F}=19$); 31 (C-6, J_{6-F}=9); (Found C, 75.59; H, 6.22; F, 3.20; C₃₄H₃₃O₅F requires C, 75.53; H, 6.19; F, 3.51%).

3.10. 1,4,5-Tri-O-benzyl-2,6-dideoxy-2-fluoro-scyllo-inositol 11

To a solution of compound **10** (140 mg, 0.26 mmol) in AcOEt (5 mL) was added a solution of sodium hydroxide (200 mg) in methanol (10 mL). The mixture was stirred for 24 h at room temperature before concentration and filtration through silica (eluent: ethyl acetate). The filtrate was concentrated under reduced pressure and used without further purification (76%). $[\alpha]_D = -2.9^{\circ}$ (*c* 0.8; CHCl₃); mp 82–85°C; ¹H NMR (300 MHz; CDCl₃) δ : 4.4 (dt, 1H, H-2, $J_{2-F}=51$; $J_{2-1}=J_{2-3}=10$); 3.5 (m, 4H, H-3, H-4, H-1, H-5); 2,4 (m, 1H, H-6eq); 1.5 (m, 1H, H-6ax); ¹³C NMR (75.50 MHz; CDCl₃) δ : 100–99 (C-2, $J_{2-F}=183$); 85 (C-4, $J_{4-F}=9$); 78 (C-3); 76 (C-5); 75 (*C*H₂Ph); 74 (C-1); 33 (C-6, $J_{6-F}=9$); (Found C, 74.30; H, 6.84; C₂₇H₂₉O₄F requires C, 74.28; H, 6.69%).

3.11. 1,4,5-Tri-O-benzyl-2,3,6-trideoxy-2,3-difluoro-*neo*-inositol 12

Diethylaminosulfur trifluoride (0.08 mL, 0.60 mmol) was added to a solution of monofluoro 11 (100 mg, 0.23 mmol) in anhydrous dichloromethane (6 mL). The mixture was stirred for 10 h under argon at room temperature before addition of methanol (10 mL). The reaction mixture was extracted and the organic layer concentrated to dryness. Flash chromatography on silica gel (eluent: AcOEt/heptane) of the residue gave the difluoro 12 (75%). $[\alpha]_{\rm D}=3.4^{\circ}$ (c 1.0; CHCl₃); mp 104–106°C; ¹H NMR (300 MHz; CDCl₃) δ : 5.0 (dd, 1H, H-3, $J_{3-F3}=51$); 4.4 (dd, 1H, H-2, $J_{2-3}=3$, $J_{2-F2}=47$); 4.0–3.8 (m, 2H, H-1, H-5); 3.4 (dd, 1H, H-4, J₃₋₄=3); 2.4 (m, 1H, H-6eq); 1.4 (m, 1H, H-6ax); ¹³C NMR (75.50 MHz; CDCl₃) δ: 95 (C-2, $J_{2-F2}=189, J_{2-F3}=24$); 92 (C-3, $J_{3-F3}=189, J_{3-F2}=24$); 80 (C-4); 76 (C-5); 75 (C-1); 32 (C-6); (Found C, 73.69; H, 6.64; C₂₇H₂₈O₃F₂ requires C, 73.95; H, 6.43%).

3.12. 2,3,6-Trideoxy-2,3-difluoro-neo-inositol 13

Compound **12** (80 mg, 0.48 mmol) dissolved in the mixture of AcOEt/MeOH (10 mL, 1:1) was hydrogenated by the same procedure described to obtain **7** from **6** (90%). $[\alpha]_D = -11.1^{\circ}$ (*c* 1.0; CH₃OH); mp 193–196°C; ¹H NMR (300 MHz; CDCl₃) δ : 4.9 (m, 1H, H-3); 4.2 (m, 1H, H-2); 3.9 (m, 1H, H-1); 3.6 (m, 1H, H-5); 3.3 (m, 1H, H-4); 2.1 (m, 1H, H-6eq); 1.2 (m, 1H, H-6ax); ¹³C NMR (75.50 MHz, CDCl₃) δ : 96 (C-2, $J_{2-F} = 168$); 93 (C-3, $J_{3-F} = 168$); 75 (C-4); 69 (C-5); 68 (C-1); 38 (C-6, $J_{6-F} = 9$).

3.13. 2,3,6-Trideoxy-2,3-difluoro-*neo*-inositol-1,4,5-tris(dibutyl)-phosphate 14

The difluoro **13** (30 mg, 0.18 mmol) was added to dibutoxy-(diisopropylamino)phosphine (300 mg, 1 mmol) and the mixture was dried for 0.5 h under reduced pressure (0.05 mmHg). Sublimated tetrazole (0.08 mg, 11.4 mmol), dissolved in dry acetonitrile (5 mL) was added under argon to the mixture. The solution was stirred under argon for 3 h and diluted with CH_2Cl_2 before *t*-BuOOH (0.21 mL, 2.1 mmol) was added. The solution was stirred under argon at room temperature for 20 h. Aqueous thiosulfate and sodium solution was added for neutralization. After extraction with dichloromethane the organic layer was purified by chromatography on silica gel (eluent: AcOEt/heptane 8:2) to yield **14** in 55% yield. $[\alpha]_D = -8.6^\circ$ (*c* 0.8; CHCl₃); ¹H NMR (300 MHz; CDCl₃) δ: 4.5–4.2 (dd, 1H, H-3, $J_{3-F} = 51$, $J_{3-4} = 12$); 4.7 (m, 1H, H-5); 4.6–4.1 (m, 3H, H-1, H-4, H-2); 4.2 (m, 12H, OCH₂CH₂CH₂CH₃); 2.8 (m, 1H, H-6eq); 1.7 (m, 13H, H-6ax, OCH₂CH₂CH₂CH₃); 1.4 (m, 12H, OCH₂CH₂CH₂CH₃); 0.9 (m, 18H, OCH₂CH₂CH₂CH₂CH₃); ¹³C NMR (75.50 MHz; CDCl₃) δ: 92 and 89 (C-3, $J_{3-F} = 180$); 91 and 88 (C-2, $J_{2-F} = 180$); 76 (C-4); 72–71 (C-1, C-5); 68 (OCH₂CH₂CH₂CH₃); 34 (C-6); 33 (OCH₂CH₂CH₂CH₃); 20 (OCH₂CH₂CH₂CH₃); 15 (OCH₂CH₂CH₂CH₃); (Found C, 48.51; H, 8.12; C₃₀H₆₁O₁₂P₃F₂ requires C, 48.38; H, 8.24%).

3.14. 3-O-Benzoyl-2,6-dideoxy-2-fluoro- *scyllo*-inositol 15

Monofluoro **10** (1.10 g, 2.03 mmol) dissolved in a mixture of AcOEt/EtOH (10 mL, 8:2) was hydrogenated for 2 h under 3 psi, in the presence of the Pd/C (800 mg). The catalyst was removed by filtration in celite and the filtrate was evaporated to dryness to afforded **15** in quantitative yield. $[\alpha]_D=11.7^{\circ}$ (*c* 1.0; CHCl₃). mp 183–185°C, ¹H NMR (300 MHz; CD₃OD) δ : 5.35 (dd, 1H, H-3, $J_{3-2}=9$, $J_{3-4}=12$); 4.5 (dt, 1H, H-2, $J_{2-F}=52$, $J_{2-1}=9$); 4.0 (m, 1H, H-4); 3.7 (m, 2H, H-1, H-5); 2.3 (m, 1H, H-6eq); 1.6 (q, 1H, H-6ax, $J_{6ax-1}=J_{6ax-5}=12$); ¹³C NMR (75.50 MHz; CD₃OD) δ : 167 (OCOPh); 97 (C-2, $J_{2-F}=184$); 76 (C-4, $J_{4-F}=9$); 75 (C-1, $J_{1-F}=18$); 70 (C-5); 68 (C-3, $J_{3-F}=18$); 37 (C-6); (Found C, 53.54; H, 5.85; C₁₃H₁₅O₅F·5/4H₂O requires C, 53.64; H, 5.94%).

3.15. 3-O-Benzoyl-2,6-dideoxy-2-fluoro-*scyllo*-inositol-1,4,5-tris(dibutyl)phosphate 16

Compound **15** (250 mg, 0.93 mmol) was phosphorylated in 73% yield by the same procedure described to obtain **14** from **13**. $[\alpha]_D = -8.6^{\circ}$ (*c* 0.8; CHCl₃); mp 44–46°C; ¹H NMR (300 MHz; CDCl₃) δ : 5.5 (t, 1H, H-3, $J_{3-4}=J_{3-2}=$ 12); 4.7 (m, 3H, H-1, H-2, H-5); 4.4 (m, 1H, H-4); 4.1 (m, 12H, OCH₂CH₂CH₂CH₃); 2.9 (m, 1H, H-6eq); 1.9 (m, 1H, H-6ax); 1.6 (12H, OCH₂CH₂CH₂CH₂CH₃); 1.4 (m, 12H, OCH₂CH₂CH₂CH₃); 0.9 (m, 18H, OCH₂CH₂CH₂CH₂CH₃); ¹³C NMR (75.50 MHz; CDCl₃) δ : 165 (Ph*C*=O); 91 (C-2, $J_{2-F}=186$); 77 (C-4); 71 (C-5, C-3); 70 (C-1); 67 (OCH₂CH₂CH₂CH₃); 32 (OCH₂CH₂CH₂CH₃); 31 (C-6); 18 (OCH₂CH₂CH₂CH₃); 13 (OCH₂CH₂CH₂CH₃); (Found C,52.48; H, 7.69; P, 10.84; F, 2.16; C₃₇H₆₆O₁₄P₃F requires C, 52.47; H, 7.85; P, 10.97; F, 2.24%).

3.16. 2,6-Dideoxy-2-fluoro-*scyllo*-inositol-1,4,5-tris(dibutyl)-phosphate 17

To a solution of monofluoro **16** (165 mg, 0.2 mmol) in dichloromethane anhydrous (12 mL) was added a solution of sodium *n*-butoxide (0.1 mol/L) in *n*-butyl alcohol. The solution was stirred for 1 h, extracted with CH₂Cl₂ and the organic layer concentrated to dryness. Flash chromatography on silica gel (eluent: AcOEt/heptane 8:2) of the residue gave the *scyllo*-inositol **17** (71%). $[\alpha]_D = -2.2^{\circ}$ (*c* 1.0; CHCl₃); mp 105–108°C; ¹H NMR (300 MHz; CDCl₃) δ : 4.8 (m, 12H, OCH₂CH₂CH₂CH₃); 4.6–4.0 (m, 4H, H-1,

H-2, H-4, H-5); 3.7 (t, 1H, H-3, $J_{3-4}=J_{3-2}=10$); 2.6 (m, 1H, H-6eq); 1.9 (m, 1H, H-6ax); 1.7 (m, 12H, OCH₂CH₂CH₂CH₃); 1.4 (m, 12H, OCH₂CH₂CH₂CH₃); 0.9 (m, 18H, OCH₂CH₂CH₂CH₂(H₃); ¹³C NMR (75.50 MHz; CDCl₃) δ : 92–90 (C-2); 74 (C-5); 72 (C-1); 70–68 (C-3, C-4); 32 (C-6); (Found C, 47.71; H, 8.19; P, 12.35; C₃₀H₆₂O₁₃P₃F requires C, 47.92; H, 8.44; P, 12.36%).

3.17. 2-O-Benzyl-6-deoxy-D-chiro-inositol 20

To a solution of *chiro*-inositol **19** (2.8 g, 6 mmol) in ethyl acetate (10 mL) was added a solution of sodium hydroxide (400 mg) in methanol (10 mL). The mixture was stirred for 4 h at room temperature before concentration and purification by chromatography on RP8 (eluent: AcOEt/MeOH) (90%). $[\alpha]_D = -2.8^{\circ}$ (*c* 1.2; CH₃OH); ¹H NMR (300 MHz; CD₃OD) δ : 4.8 (m, 2H, OCH₂Ph); 4.0 (m, 2H, H-1, H-3); 3.6 (m, 2H, H-2, H-5); 3.5 (m, 1H, H-4); 2.8 (m, 2H, H-6eq, H-6ax); ¹³C NMR(75.50 MHz; CD₃OD) δ : 80 (C-2); 74 (C-4); 70 (C-3); 68 (C-5); 66 (C-1); 38 (C-6); (Found C, 60.16; H, 7.17; C₁₃H₁₈O₅·1/3H₂O requires C, 60.33; H, 7.23%).

3.18. 2-*O*-Benzyl-6-deoxy-D-*chiro*-inositol-1,3,4,5-tetra(dibutyl)-phosphate 21

The tetrol 20 (100 mg, 0.4 mmol) was dissolved in THF (7 mL) and cooled to 0°C under argon. n-Butyllithium 1.6 M solution in hexane was added (0.1 mL, 1.0 mmol). The temperature was cooled to -40° C and di-*n*-butyl phosphoryl chloride (0.72 g, 3.14 mmol) was added. The mixture was stirred for 1 h extracted with CH₂Cl₂ and the organic layer concentrated to dryness. The product was separated by chromatography on silica gel (eluent: AcOEt/heptane 8:2) (30%). $[\alpha]_{D} = -13.2^{\circ}$ (*c* 1.0; CH₃OH). ¹H NMR (300 MHz; CDCl₃) δ: 4.8-4.4 (m, 5H, H-1, H-3, H-2, H-5, H-4); 3.9 (m, 16H, OCH₂CH₂CH₂CH₃); 2.5-2.0 (m, 2H, H-6eq, H-6ax); 1.5 (m, 16H, OCH₂CH₂CH₂CH₃); 1.2 (m, 16H, OCH₂CH₂CH₂CH₃CH₃); 0.7 (m, OCH₂CH₂CH₂CH₂CH₃); 13 C NMR (75.50 MHz; CDCl₃) δ : 74 (C-1); 73 (C-3); 72 (C-2); 71 (C-5, C-4); 66 (OCH₂CH₂CH₂CH₃); 31 (OCH₂CH₂CH₂CH₃); 30 (C-6); 19 (OCH₂CH₂CH₂CH₂CH₃); 14 (OCH₂CH₂CH₂CH₃); (Found C, 53.21; H, 8.73; C₄₅H₈₆O₁₇P₄ requires C, 52.83; H, 8.47%).

3.19. 6-Deoxy-D-*chiro*-inositol-1,3,4,5-tetra(dibutyl)phosphate 22

The tetraphosphate 21 (100 mg, 0.1 mmol) dissolved in AcOEt/EtOH (10 mL, 3:1) was hydrogenated for 3 h under 3 psi, in the presence of the Pd/C catalyst (100 mg). The catalyst was removed by filtration in silica (eluent: AcOEt) and the solvent evaporated to obtain 22 in quantitative yield. $[\alpha]_D = -17^\circ$ (c 1.9; CHCl₃); ¹H NMR (300 MHz; CDCl₃) δ: 4.9 (m, 1H, H-4); 4.6 (m, 3H, H-2, H-1, H-5); 4.3 (m, 1H, H-3); 4.1 (m, 16H, OCH₂CH₂CH₂CH₃); 3.6 (m, 1H, OH); 2.4 (m, 1H, H-6eq); 2.3 (m, 1H, H-6ax); 1,7 (m, 16H, $OCH_2CH_2CH_2CH_3$); $(m, 24H, OCH_2CH_2CH_2CH_3); 1.0 (m, 24H,)$ 1.3 OCH₂CH₂CH₂CH₂CH₃); 13 C NMR (75.50 MHz; CDCl₃) δ : 75 (C-4, C-3); 72 (C-2); 69 (C-1, C-5); 68 (OCH₂CH₂CH₂CH₃); 32 (OCH₂CH₂CH₂CH₃); 30 (C-6); 19 (OCH₂CH₂CH₂CH₃); 14 (OCH₂CH₂CH₂CH₃); (Found C, 49.01; H, 8.59; P, 13.32; C₃₈H₈₀O₁₇P₄ requires C, 48.92; H, 8.64; P, 13.28%).

Acknowledgements

This research was supported by CNPq and Bayer-AG Geschaftsbereich Pharma und Entwicklung Istitut. We thank Dr. Jeannine Cleophax for many helpful discussions.

References

 (a) Reitz, H. L. In Inositol Phosphates Derivatives, In ACS Symposium Series, ACS: Washington, DC, 1991; pp 1.
 (b) Potter, B. V. L. Nat. Proc. Rep. 1990, 7, 1. (c) Billington, D. C. Chem. Soc. Ver. 1989, 18, 83. (d) Lapetina, D. C. Phosphoinositides and Receptor Mechanisms 1986, 7, 271.
 (e) Potter, B. V. L.; Lampe, D. Angew. Chem., Int. Ed. Engl. 1995, 34, 1933. (f) Erneux, C.; Takazawa, K. Trends Pharm. Sci. 1991, 12, 1745. (g) Powis, C. Trends Pharm. Sci. 1992, 12, 188. (h) Powis, C.; Aksoy, I. A.; Melder, D. C.; Aksoy, S.; Eichinger, H.; Fauq, A.; Kozikowski, A. P. Cancer Chemother. Pharmacol. 1991, 29, 95. (i) Billington, D. C. The Inositol Phosphates, Chemical Synthesis and Biological Significance, V.C.H.: Weinheim, 1993 and references therein.

- (a) Lûckhoff, A.; Clapham, D. E. *Nature (London)* **1992**, *355*, 356.
 (b) Wilcox, R. A.; Challis, R. A. J.; Baudin, G.; Vasella, A.; Potter, B. V. L.; Nahorski, S. R. *Biochem. J.* **1993**, *294*, 191.
- Vieira de Almeida, M.; Dubreuil, D.; Cleophax, J.; Verre-Sebrié, C.; Pipelier, M.; Prestat, G.; Vass, G.; Gero, S. D. *Tetrahedron* 1999, 55, 7251 and references therein.
- (a) Dahl, J.; Jurczak, A.; Cheng, L. A.; Baker, D. C.; Benjamin, T. L. J. Virol. **1998**, 72, 3221. (b) Kozikowski, A. P.; Fauq, A. H.; Powis, G.; Melder, D. C. J. Am. Chem. Soc. **1990**, 112, 4528.
- Safrany, S. T.; Wojcikiewicz, R. J. H.; Strupish, J.; Nahorski, S. R.; Dubreuil, D.; Cleophax, J.; Gero, S. D.; Potter, B. V. L. *FEBS Lett.* 1991, 278, 252.
- Cleophax, J.; Dubrueil, D.; Gero, S. D.; Loupy, A.; Vieira de Almeida, M.; Da Silva, A. D.; Vass, G. *Bioorg. Med. Chem. Lett.* 1995, 5, 831.
- (a) Lowe, G.; McPhee, F. J. Chem. Soc. Perkin Trans. 1 1991, 1249. (b) Sawyer, D. A.; Potter, B. V. L. J. Chem. Soc. Perkin Trans. 1 1992, 923. (c) Moyer, J. D.; Reizes, O.; Ahir, S.; Jiang, C.; Malinowski, N.; Baker, D. C. Mol. Pharmacol. 1989, 33, 683.
- Dubreuil, D.; Cleophax, J.; Vieira de Almeida, M.; Verre-Sebrié, C.; Liaigre, J.; Vass, G.; Gero, S. D. *Tetrahedron* 1997, 53, 16747.
- Da Silva, A. D.; Benício, A. A. A.; Gero, S. D. Tetrahedron Lett. 1999, 40, 6531.